This article was downloaded by: On: 23 January 2011 Access details: Access Details: Free Access Publisher Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37- 41 Mortimer Street, London W1T 3JH, UK

Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information: <http://www.informaworld.com/smpp/title~content=t713617200>

Synthesis of Acetylated Ranunculin Diastereoisomers and δ-Glucosyloxyγ-Oxo Esters from α or β Glucosylmethylfurfural

Louis Cottier^a; Gérard Descotes^a; Yaya Soro^a a École Supérieure de Chimie Physique Electronique de Lyon, Laboratoire de Chimie Organique 2, UMR CNRS 5181, Université Claude Bernard Lyon 1, Villeurbanne Cedex, France

To cite this Article Cottier, Louis , Descotes, Gérard and Soro, Yaya(2005) 'Synthesis of Acetylated Ranunculin Diastereoisomers and δ-Glucosyloxy-γ-Oxo Esters from α or β Glucosylmethylfurfural', Journal of Carbohydrate Chemistry, 24: 1, $55 - 71$

To link to this Article: DOI: 10.1081/CAR-200049688 URL: <http://dx.doi.org/10.1081/CAR-200049688>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use:<http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

Journal of Carbohydrate Chemistry, 24:55–71, 2005 Copyright \odot Taylor & Francis, Inc. ISSN: 0732-8303 print DOI: 10.1081/CAR-200049688

Synthesis of Acetylated Ranunculin Diastereoisomers and δ -Glucosyloxy- γ -Oxo Esters from α or β **Glucosylmethylfurfural**

Louis Cottier, Gérard Descotes, and Yaya Soro

École Supérieure de Chimie Physique Electronique de Lyon, Laboratoire de Chimie Organique 2, UMR CNRS 5181, Université Claude Bernard Lyon 1, Villeurbanne Cedex, France

The α and β acetylated (5-formyl-2-furyl)methyl-D-glucopyranosides were synthesized and converted into acetylated ranunculin diastereoisomers and δ -glucosyloxy- γ -oxo esters.

Keywords Synthesis, Glycosylation, Furanic compounds, Photooxygenation

INTRODUCTION

Ten years ago, we were interested in the synthesis of 5-hydroxymethylfurfural (HMF) obtained from fructose or disaccharides^[1,2] and its conversion into hydroxybutenolides, γ -oxo acids, γ -oxo esters, and 5-aminolevulinic acid hydrochlororide.^[3,4] More recently, we applied our methodology^[3] to the synthesis of the diastereoisomers 1α and 2β of [(2S)-5-oxo-2,5-dihydro-2-furanyl]methyl 2,3,4,6-tetra-O-acetyl- β -D-glucopyranoside (ranunculin tetraacetate) [2 β (S)] (Schs. 1 and 3). Ranunculin tetracetate is a precursor of $(-)$ -ranunculin (3β))

Received February 2, 2004; accepted November 29, 2004.

Address correspondence to Louis Cottier, CPE, Bt 308, Laboratoire de Chimie Organique 2, , Université Claude Bernard Lyon 1, 43 Boulevard du 11 Novembre 1918, 69622 Villeurbanne Cedex, France. Fax: 33 472 4481 60; E-mail: louis.cottier@univ-lyon1.fr

Scheme 1

having cytotoxic and antimutagenic activities.^[5,6] We also extended our methodology to the preparation of the δ -glucosyloxy- γ -oxo esters 4α and 5β , precursors of GABA derivative 6 bearing a glucosyloxymethyl moiety (Sch. 3). This work is in line with the recent publications of Lichtenthaler et al.^[7,8] on the synthesis of heterocycles from carbohydrate feedstocks. The structure of $(-)$ ranunculin (3B) was established to be a β -glucoside of a y-hydroxymethyl- α , β -butenolide.^[9–11] The first total synthesis of 3β was described by Cardellach et al.[12,13] using a Koenigs-Knorr reaction between (5S)-5-(hydroxymethyl)- $2(5H)$ -furanone (7) and the tetra-O-acetyl- α -D-glucopyranosyl bromide (8a) in the presence of silver oxide (Ag_2O) , followed by acidic hydrolysis. In the first

publication,^[12] the peracetate of ranunculin 2β was obtained as a mixture of epimers, while in a second publication,^[13] a compound 3β identical with the natural product was synthesized (Sch. 1). However, a large excess of both $Ag₂O$ and 8a was used, which may appear somewhat limiting.

Some years later, Fang et al.^[14] published the preparation of $(-)$ -ranunculin 3β from mannitol in six steps and a rather low 15% overall yield. Moreover,

the α -anomer of the per-O-benzoyl ranunculin derivative was obtained from isomaltulos $e^{[15]}$ in four steps. Indeed, the oxidative degradation of isomaltulose in alkaline medium, followed by product conversion into lactones, subsequent benzoylation, and samarium diiodide mediated elimination, led to the per-Obenzoylester with an overall yield of 36%. In this paper, we describe the preparation of acetylated ranunculin diastereoisomers 1α and 2β from the furanyl glucopyranosides 9α or 10β (α and β GMF), respectively (Sch. 3), via the photooxygenation of the furanic ring. This new approach to the various diastereoisomeric targets should help the study of their biologic activities.

RESULTS AND DISCUSSION

The acetylated GMF (9α) was obtained from isomaltulose (11) according to Lichtenthaler's procedure^[15] in 48% yield after peracetylation (Sch. 2). However, the preparation of the corresponding anomer 10β via the condensation between a glycosyl donor 8a–c and an alcohol furanic acceptor 12a–d was more difficult (Table 1).

The first coupling attempt of **8a** and HMF (12a) in the presence of Ag_2O led to 10β in poor yield, probably due to the electron withdrawing effect of the aldehyde moiety decreasing the nucleophilicity of the hydroxyl group (Sch. 2). The transformation of the aldehyde function into an acetal moiety, to give 12b, did not improve the yield. The substitution of HMF by furfuryl alcohol (12c) led to the glucoside 13β in a higher yield (80%). The latter was easily converted into 10β by a Vilsmeier-Haack formylation (74% yield). Two other alternatives to prepare 10β were also studied. The method of Schmidt^[16] using imidate 8b and HMF (12a) gave 10 β in moderate yield (32%). Analogously, the reaction between glucose pentaacetate (8c) and the

Compounds			(%) Products	
8	12	Experimental conditions ^{σ}	10 $\boldsymbol{\beta}$	13B
o	α	$Ag2O$, rt		
a	b	Ag ₂ O, I_2 , CaSO ₄ , rt	8	
a	c	Ag ₂ O, I_2 , CaSO ₄ , rt		80
b	$\mathbf{a}^{\scriptscriptstyle D}$	$BF_3.Et_2O$, $-20^{\circ}C$ then rt	32	
b	d	$BF_3.Et_2O$, -20° C then rt		
с	c	$BF_3.Et_2O$, -20° C then rt	trace	
с	d	$BF_3.Et_2O$, $-20^{\circ}C$ then rt	33	

Table 1: Condensation between glucose derivatives 8 and the furanic alcohols 12.

 σ Solvent: CH₂Cl₂.
bSolvent CH₃CN.

silylated HMF derivative 12d according to Nair and Joseph^[17] yielded 10 β in a similar yield. Since glucose pentaacetate is cheaper and more stable than the corresponding glucosyl bromide, and since the silylated 12d is more stable than HMF 12a, the latter strategy appeared more convenient.

The photooxygenation of GMF 9α with singlet oxygen^[3] led to the corresponding hydroxy- Δ^2 -butenolide 14 α as a 1/1 diastereoisomeric mixture (Sch. 3). These compounds are very unstable and easily decomposed on column chromatography.

Due to their poor stability during column chromatography, the $1/1$ mixture of diastereoisomers 14α was acetylated (acetic anhydride and sodium acetate)^[18] to give the corresponding isomers 16α , which were not separated. However, NMR data of the crude mixture of 16α showed the presence of a major epimer (70%). The increased proportion of one epimer was attributed to the kinetically esterification of the corresponding hydroxy- Δ^2 -butenolide diastereoisomer 14 α . Under identical conditions, the acetylated isomers 17 β were prepared as a $54/46$ R/S mixture, from 10β . In this case, the diastereoisomers were separated by chromatography.

As a hemiacetalic cyclic compound is in equilibrium with the alicyclic tautomeric form, the compounds 14α or 15β were reduced to the corresponding acetylated $(5'-oxo-2', 5'-dihydrofuran-2'-yl)$ methyl- α or β -D-glucopyranoside (1α) or (2β) . Unfortunately, the sugar moiety did not induce any stereoselectivity (Table 2, entry 1). Even the addition of ceric chloride, which, according to Kumar et al.,^[19] favors the stereospecific reduction of 7-oxocholesterylacetate, did not improve significantly the stereoselectivity, whatever the temperature used (Table 2, entries 2 and 3). Besides, attempted reduction with 9-BBN failed (Table 2, entry 4).

An X-ray analysis of first eluted epimer 17β , showing an R configuration for carbon C-2', allowed the possibility to assign the configuration R or S at this carbon for each of the epimers 17β . The synthesis of pure $2\beta(S)$, identical

		Compounds, products		
Entry	Experimental conditions $^{\circ}$	14 α . 1 α (Yields %, R/S %)	15 β , 2 β (Yields %, R/S %)	
	N aBH ₄ , MeOH, rt	80, 50/50	81, 50/50	
2	NaBH ₄ , CeCl ₃ , 7H ₂ O (0.4M) MeOH, rt	80, 40/60	82, 60/40	
3	NaBH ₄ , CeCl ₃ , 7H ₂ O (0.4M) MeOH, -78° C then rt	80, 40/60	80, 60/40	
$\boldsymbol{\varDelta}$	9-BBN, THE, 0° C then rt	Trace	Trace	

Table 2: Reduction of hemiacetals 14α and 15β .

 α After the total reduction, the pH of the solution was adjusted to 1 with HCl (5N).

to $(-)$ ranunculin tetraacetate according to Camps et al.^[13] and its epimerisation into $1\alpha(S)$ with TiCl₄ according to Dasgupta et al.,^[20] allowed to assign the chemical shift of protons and carbons of the other stereoisomers $1\alpha(R)$ and $2\beta(R)$, and to estimate the epimers ratio. The R/S configuration of carbon C-2' in epimers 16α was attributed by comparing their NMR spectra and those of 17β , 1α (R or S) and 2β (R or S).

The hydroxy- Δ^2 -butenolides ${\bf 14}\alpha$ and ${\bf 15}\pmb{\beta}$ were easily transformed into the corresponding γ -oxo esters 4α and 5β respectively, via the corresponding γ -oxo acids (Sch. 3). Some years ago, γ -oxo acids and γ -oxo esters were obtained by photooxygenation of furanic compounds and reduction under ultrasound with Zn in acetic acid.^[3] As Misiti et al.^[21] described the chemoselective catalytic hydrogenation of α, β unsaturated esters using Pd/C, the isomers 14α and 15β were selectively reduced via a hydrogen transfer reaction in the presence of the potassium formate/palladium(II) acetate system.^[22] The γ -keto acids intermediates were not isolated but transformed into γ -keto esters 4α and 5β .

CONCLUSION

Several methods of synthesis of α - and β -GMF 9α and 10β were compared. If the synthesis of α -GMF 9α is easy from isomaltulose, the preparation of β -GMF 10 β is more difficult. The best yield was obtained via the condensation of tetra-O-acetyl- α -D-glucopyranosyl bromide (8a) with furfuryl alcohol (12c) followed by a formylation reaction. The direct condensation of HMF (12a) with a glucosyl donor gave a moderate yield $(32%)$ of 10β . The photooxygenation of the furanic moiety of α and β GMF, and subsequent reduction, led to the acetylated diastereoisomers of ranunculin 1α and 2β in moderate yield. However, this methodology gave the possibility to prepare both acetylated epimers 1α and 2β of α and β ranunculin in two steps from α and β GMF. The selective reduction of the intermediate hydroxy butenolides 14α and **15** β , via a hydrogen transfer reaction, gave access to γ -keto esters 4α and **56**, precursors of GABA derivatives with a glycosyl moiety having an α or β configuration.

EXPERIMENTAL

Isomaltulose was kindly provided by Südzucker AG Company (Germany). Melting points were determined on a Buchi melting point apparatus and are uncorrected. Optical rotations were measured with a Perkin Elmer Model 241 polarimeter. The 200 MHz or 300 MHz ¹H-NMR and 50 or 75 MHz 13 C-NMR were recorded with a Bruker AC 200 or AM 300 spectrometer with TMS as an internal standard. In the ¹H- and ¹³C-NMR spectra, the chemical shift of the protons or the carbons of each enantiomer 14α and 15β was

assigned using the COSY and HSQC procedures. Mass Spectra were recorded with a Finnigan Mat 95 XL spectrometer. Thin layer chromatography (TLC) was carried out on plates coated with silica gel 60. Column chromatography was carried out on silica gel Si 60; the ratio of solvents were measured in volume.

(5'S) (5'-Oxo-2',5'-dihydrofuran-2'-yl)methyl-2,3,4,6-tetra-Oacetyl- β -D-glucopyranoside (2 β)

The compound $2\beta(S)$ was prepared according to Camps et al.^[13] 0.5 g (4.38 mmol) of butenolide 7 in dichloromethane with silver oxide $(3 \times 1.12 \text{ g})$, 4.8 mmol) and bromide 8a $(3 \times 1.93 \text{ g}, 4.38 \text{ mmol})$ led to $2\beta(S)$ (0.44 g, 1.0 mmol) in 23% yield after purifying by column chromatography (dichloromethane/ethyl acetate: 70/30). **2** β (S): white solid; mp 135–136°C lit^[13] mp 136–138°C; $[\alpha]_D^{20}$ –22.6 (c 1.1, CH_2Cl_2) $\text{lit}^{[13]}$ $[\alpha]_D^{20}$ –24.2 (c 1.1, CHCl₃); ¹H-NMR (200 MHz, CDCl₃): δ 2.02-2.09 (4s, 12H, -OCOCH₃), 3.70 (m, 1H, H-5) , 3.80 (dd, 1H, J_{CH_a} , $\text{H}_{2'} = 4.8$, $J_{\text{gemCH}_2} = 11.0 \text{ Hz}$, -OCH₂), 4.08 (dd, 1H, J_{CH_b} , $H'_2 = 4.80$, $J_{\text{gemCH}_2} = 11.0 \text{ Hz}$, $-OCH_2$), 4.12 (dd, 1H, $J_{5,6a} = 2.2$, $J_{6a,6b} = 12.4 \text{ Hz}, \text{ H-6}_a$, 4.23 (dd, 1H, $J_{5,6b} = 4.5, J_{6a,6b} = 12.4 \text{ Hz}, \text{ H-6}_b$), 4.60 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1), 5.08 (dd, 1H, $J_{1,2} = 7.9$, $J_{2,3} = 9.2$ Hz, H-2), 5.15 (t, 1H, $J_{3,4} = J_{4,5} = 9.2 \text{ Hz}$, H-4), 5.15 (m, 1H, H-2'), 5.22 (t, H, $J_{2,3} = J_{3,4} = 9.2 \,\text{Hz}$, H-3), 6.20 (dd, 1H, $J_{2',4'} = 2.0$, $J_{3',4'} = 5.7 \,\text{Hz}$, H-4'), 7.47 (dd, 1H, $J_{2',3'} = 1.5$, $J_{3',4'} = 5.7$ Hz, H-3') similar to literature data;^[13] ¹³C-NMR (75 MHz, CDCl₃): δ 20.6–20.7 (4 CH₃), 61.7 (C-6), 68.3 (C-4), 69.3 $(CH₂), 70.8 (C-2), 72.0 (C-5), 72.5 (C-3), 81.4 (C-2'), 100.9 (C-1), 122.8 (C-4'),$ 153.0 (C-3'), $169.5-170.1$ (4 CO₂), 172.4 (C-5').

[5-(Dimethoxymethyl)-2-furyl]methanol (12b)

To a solution of 12a $(0.3 g, 2.4 mmol)$ in methanol $(7 mL)$ maintained at 0° C were added 0.1 g (0.23 mmol of Ytterbium) of Ytterbium sulfate supported on Amberlite $15^{[23]}$ and trimethyl orthoformate $(0.3 \text{ mL}, 2.78 \text{ mmol})$. The mixture was allowed to warm to rt and stirred for 2 hr. Then trimethyl orthoformate (1 mL, 9.15 mmol) was added and the solution was stirred for 2 hr more. The solution was filtered using a glass apparatus previously washed with alcaline solution. Volatiles were evaporated under vacuum. $12b(0.33g)$, 1.92 mmol, 80%) was obtained pure as a yellow oil. $12b:$ ¹H-NMR (200 MHz, CDCl₃): δ 2.88 (s broad, 1H, OH), 3.33 (s, 6H, $-OCH_3$), 4.56 (s, 2H, $-OCH_2$), 5.38 (s, 1H, CH(OCH₃)₂), 6.24 (d, 1H, $J_{3,4} = 3.1$ Hz, H-3), 6.34 (d, 1H, $J_{3,4} = 3.1$ Hz, H-4); ¹³C-NMR (75 MHz, CDCl₃): δ 52.8, 57.8 (2 – OCH₃), 66.3 $(-OCH₂), 109.1 (C-3), 109.7 (C-4), 151.1 (C-2), 151.8 (C-5).$

Anal. Calcd for $C_8H_{12}O_4$ (172.18): C, 55.80; H, 7.02. Found: C, 55.67; H, 6.99.

5-({[Tert-butyl(dimethyl)silyl]oxy}methyl)-2-furaldehyde (12d)

To a solution of $12a$ (6.3 g, 50 mmol) in dimethylformamide (12.6 mL) were added imidazole (8.5 g, 125 mmol) and ter-butyldimethylsilyl chloride (8.9 g, 59 mmol). The solution was stirred for 24 hr. The silylated compound was extracted with petroleum ether $(5 \times 25 \text{ mL})$. After washing with brine, drying, and evaporating the solvent, the remaining oil was distillated $(132^{\circ}C/1.5$ Torr) giving 12d $(9g, 3.7$ mmol, 74%) as a light yellow liquid. 12d: ¹H-NMR: similar to literature data.^[24]

(5'-Formyl-2'-furyl)methyl-2,3,4,6-tetra-O-acetyl- α -Dglucopyranoside (9 α)

To a solution of isomaltulose monohydrate 11 (4.5 g, 11.5 mmol) in dimethyl sulfoxide (45 mL), heated to 120 $^{\circ}$ C, was added 0.4 g of strongly acidic sulfonic acid-type ion-exchange resin (Dowex 50-WX4, H^+ form, dried) and stirred at 120° C for 7 hr. After filtration, the filtrate was removed under reduced pressure at 80° C and the residual brown syrup was dissolved in water (25 mL). This solution was extracted with dichloromethane (2×10 mL). The aq. phase was evaporated under reduced pressure giving an oil (7.6 g), which was dissolved in pyridine (380 mL). After adding acetic anhydride, this new solution was stirred for 24 hr. Cold water (150 mL) was added, and the mixture was treated with sulfuric acid $(2N, 1mL)$, neutralized with aq. sodium hydrogenocarbonate (5 mL), washed with water, and dried. After filtration and evaporation, the raw material was purified by column chromatography (petroleum ether/ethyl acetate: $50/50$) giving 9α (2.75 g, 6 mmol, 48%) as a yellow oil. 9α : [α] $_D^{20}$ +132 (c 1.7, CHCl₃) lit.^[15] +133 (c 1.7, CHCl₃); ¹H-NMR and ¹³C-NMR: similar to literature data.^[15]

(5'-Formyl-2'-furyl)methyl-2,3,4,6-tetra-O-acetyl-ß-Dglucopyranoside (10 β)

(a) Koenigs-Knorr method: To a solution of HMF 12a (0.9 g, 2.2 mmol) in dry dichloromethane (15 mL) were added $8a (0.9 g, 2.2 mmol)$ and silver oxide (0.41 g, 17.6 mmol). After stirring for 5 hr at rt, the mixture was filtered and the solvent evaporated. The residue was chromatographed through silica gel (dichloromethane/ethyl acetate: $80/20$) giving pure product 10β (0.11 g, 0.24 mmol, 11%) as a yellow solid. 10 β : mp 130°C; [a] $^{20}_{\text{D}}$ –39 (c 1, CH₂Cl₂); 1 H-NMR (200 MHz, CDCl₃): δ 2.03 – 2.09 (4s, 12H, -OCOCH₃), 3.75 (m, 1H, H-5), 4.18 (dd, 1H, $J_{5,6a} = 3.2$, $J_{6a,6b} = 12.8$ Hz, H-6_a), 4.25 (dd, 1H, $J_{5,6b} = 6.5, J_{6a,6b} = 12.8 \text{ Hz}, H-6_b$, 4.62 (d, 1H, $J_{1,2} = 7.8 \text{ Hz}, H-1$), 4.70 (d, 1H, $J_{\text{gem CH}_2} = 13.7 \text{ Hz}$, CH₂), 4.84 (d, 1H, $J_{\text{gem CH}_2} = 13.7 \text{ Hz}$, CH₂), 5.00 (dd, 1H, $J_{1,2} = 7.8$, $J_{2,3} = 9.2$ Hz, H–2), 5.10 (dd, 1H, $J_{3,4} = J_{4,5} = 9.2$ Hz, H– 4), 5.20 (dd, 1H, $J_{2,3} = J_{3,4} = 9.2$ Hz, H-3), 6.53 (d, 1H, $J_{3',4'} = 3.5$ Hz, H-3'), 7.22 (d, 1H, $J_{3',4'} = 3.5$ Hz, H-4'), 9.63 (s, 1H, CHO); ¹³C-NMR (75 MHz, CDCl₃): δ 20.7–20.9 (4 CH₃), 61.8 (C–6), 68.2 (C–4), 71.0 (C–2), 72.0 (C–5), 72.6 (C-3), 99.8 (C-1), 111.8 (C-3'), 122.1 (C-4'), 152.8 (C-5'), 156.7 (C-2'), 169.5–170.0 (4 COCH3),177.6 (CHO).

Anal. Calcd for $C_{20}H_{24}O_{12}$ (456): C, 52.63; H, 5.30. Found: C, 52.42; H, 5.16. (b) According to Schmidt et al. [16]: Under argon, $12a(0.14g, 1.1 \text{ mmol})$ and 8b (1.08 g, 2.2 mmol) were dissolved in a mixture of acetonitrile (22 mL) and dry dichloromethane (10 mL). After addition of molecular sieve (1 g), boron trifluoride diethyl etherate (0.6 mL, 4.4 mmol) was added to the solution, cooled to 0° C. The mixture was stirred for 2 hr at rt, and then diluted with dichloromethane (10 mL), neutralized with sodium hydrogenocarbonate, washed with water, and dried. After filtration, the solvent was evaporated. The residue was chromatographed through column silica gel (dichloromethane/ethyl acetate: 80/ 20) giving pure 10β (0.16 g, 0.35 mmol, 32%).

(c) According to Nair and Joseph:^[17] The compound $12d$ (0.25 g, 2 mmol) then boron trifluoride diethyl etherate (4 mL) were added at -20° C to a solution of $\&c\ (0.86\,g, 2.2\,\text{mmol})$ in dry dichloromethane $(15\,\text{mL})$. After stirring for 5 hr at -20° C, the mixture was neutralized with sodium hydrogenocarbonate and diluted with diethyl ether (75 mL). The organic phase was washed with water $(2 \times 15 \text{ mL})$ and then brine (10 mL), and dried. After filtration, the solvent was evaporated. The raw product was purified by column chromatography (dichloromethane/ethyl acetate: $80/20$) giving pure 10β (0.302 g, 0.66 mmol, 33%).

(d) From 13β : A solution of phosphorus oxychloride in dimethylformamide (0.7 mL) was previously prepared by adding phosphorus oxychloride (0.9 mL, 9.2 mmol) in dimethylformamide maintained below 20° C. This orange solution, cooled to 0° C, was dropped in a cooled solution of 13β (1.96 g, 4.5 mmol) in 1,2-dichloroethane (4.5 mL). This mixture was allowed to warm to rt, stirred for 8 hr, and then neutralized with an aq. solution of sodium acetate $(3.1 g \text{ in } 14 \text{ mL})$ and diluted with diethyl ether (80 mL) . The organic solution was washed with brine $(2 \times 10 \,\text{mL})$, dried, and evaporated. The crude product was purified by column chromatographed with an elution gradient (dichloromethane then dichloromethane/ethyl acetate until, 80/20) giving 10β (1.52 g, 3.3 mmol, 74%).

2'-Furylmethyl-2,3,4,6-tetra-O-acetyl-ß-D-glucopyranoside (13β)

To a solution of $12c$ (9.6 g, 0.1 mmol) in dichloromethane (40 mL) were added dry calcium sulfate $(4g)$ and silver carbonate $(5.5g, 20 \text{ mmol})$. After stirring for 30 min, iodine $(1.52 g, 6.2 mmol)$ was added and then was

dropped 8a $(8.22 g, 20 mmol)$ dissolved in dichloromethane $(30 mL)$. The mixture was stirred at room temperature for 3 hr and then centrifugated to eliminate the mineral salts. The organic phase was washed with sodium thiosulfate $(2 \times 10 \text{ mL})$ and with brine, and dried. The solvent was evaporated, and the raw solid was recrystallized (petroleum ether/ethyl acetate) giving 13β pure (6.9 g, 15.6 mmol, 80%). ${\bf 13\beta}$: mp ${\bf 110{-}111}^{\circ}\rm C;$ $[\alpha]_{\rm D}^{20}$ $+{\bf 47}$ (c 1, $\rm CHCl_3);$ $^1\rm H$ -NMR (200 MHz, CDCl₃): δ 2.00–2.10 (4s, 12H, -OCOCH₃), 3.68 (ddd, 1H, $J_{4,5} = 9.7, J_{5,6a} = 2.5, J_{5,6b} = 4.6 \text{ Hz}, H_{5,6b} = 4.13 \text{ (dd)}, H_{5,6a} = 2.5,$ $J_{6a,6b} = 12.3 \text{ Hz}, \text{ H}-6a$, 4.26 (dd, 1H, $J_{5,6b} = 2.5, J_{6a,6b} = 4.6 \text{ Hz}, \text{ H}-6b$), 4.56 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1) 4.61 (d, 1H, $J_{\text{gem CH}_{2}} = 13.2$ Hz, CH₂), 4.74 (d, 1H, $J_{\text{gem CH}_2} = 13.2 \text{ Hz}, \text{ CH}_2$), 4.98 (dd, 1H, $J_{1,2} = 7.9, J_{2,3} = 9.3 \text{ Hz}, \text{ H}-2$), 5.08 (dd, 1H, $J_{3,4} = J_{4,5} = 9.3$ Hz, H-4), 5.17 (dd, 1H, $J_{2,3} = J_{3,4} = 9.3$ Hz, H-3), 6.33 (dd, 1H, $J_{3',4'} = 3.3$, $J_{3',5'} = 0.8$ Hz, H-3'), 6.35 (dd, 1H, $J_{3',4'} = 3.3$, $J_{4',5'} = 1.8 \,\text{Hz}, \text{ H-4'}, \text{ 7.40 } \text{ (dd, 1H, } J_{3',5'} = 0.8, \text{ } J_{4',5'} = 1.8 \,\text{Hz}, \text{ H-5'});$ ¹³C-NMR (75 MHz, CDCl₃): δ 20.5–20.8 (4 CH₃), 62.2 (C–6), 63.0 (CH₂), 68.8 $(C-4)$, 71.4 $(C-2)$, 72.2 $(C-5)$, 73.1 $(C-3)$, 99.3 $(C-1)$, 110.6 $(C-3')$, 110.8 $(C-4')$, 143.0 $(C-5')$, 150.6 $(C-2')$, 169.3-170.6 (4 COCH₃).

Anal. Calcd for $C_{19}H_{24}O_{11}$ (428.38): C, 53.27; H, 5.65. Found: C, 53.23; H, 5.63.

(2'-Hydroxy-5'-oxo-2',5'-dihydrofuran-2'-yl)methyl-2,3,4,6tetra-O-acetyl- α -D-glucopyranoside (14 α R+S)

Rose Bengal-Sephadex resin^[25] (109 mg) was added to 9α (302 mg, 0.66 mmol) dissolved in a mixture of dichloromethane (4 mL) and ethyl alcohol (9 mL). The mixture, under a low pressure of dioxygen, was quickly stirred and irradiated for 6 hr with a halogen lamp (150 W). After filtration and evaporation of the solvents, the residue was chromatographed on a preparative TLC plate (dichloromethane/ethyl acetate: $50/50$) giving 14α $(0.261 \,\mathrm{mg},\, 0.57 \,\mathrm{mmol},\, 86\%)$ as a mixture of epimers $(R/S: 1/1)$. A second preparative plate chromatography with a little amount led to two fractions I and II having one of both epimers in a bigger amount. 14α (R+S), solid; mp $45-46^{\circ}$ C; fraction I: ¹H-NMR (200 MHz, CDCl₃): $\delta 2.00-2.10$ (4s, 12H, $-OCOCH_3$), 2.20 (s, 1H, OH); 3.70 (d, 1H, $J_{\text{gem CH}_2} = 11.0 \text{ Hz}$, CH₂), 3.90 (d, 1H, $J_{\text{gem CH}_2} = 11.0 \text{ Hz}$, CH₂)_, 4.05 (m, 2H, H-5, H-6_b), 4.25 (dd, 1H, $J_{5,6a} = 4.4, J_{6a,6b} = 12.5 \text{ Hz}, \text{ H}-6_a$, 4.86 (dd, 1H, $J_{1,2} = 3.7, J_{2,3} = 9.6 \text{ Hz}, \text{ H}-$ 2), 5.01 (dd, 1H, $J_{3,4} = J_{4,5} = 9.6$ Hz, H–4), 5.10 (d, 1H, $J_{1,2} = 3.7$ Hz, H–1), 5.41 (dd, 1H, $J_{2,3} = J_{3,4} = 9.6$ Hz, H-3), 6.19 (d, 1H, $J_{3',4'} = 5.6$ Hz, H-4'), 7.22 (d, 1H, $J_{3',4'} = 5.6$ Hz, H-3'); ¹³C-NMR (75 MHz, CDCl₃): δ 20.6–20.7 (4 CH₃), 61.8 (C–6), 67.8 (CH₂), 68.0 (C–5), 68.4 (C–4), 70.1 (C–3), 70.4 (C–2), 96.9 $(C-1)$, 105.6 $(C-2')$, 125.2 $(C-4')$, 147.9 $(C-3')$), 169.2-170.8 (4 COCH₃), 170.1 (C-5'); fraction II: ¹H-NMR (200 MHz, CDCl₃): δ 2.00-2.10 (4s, 12H, $-OCOCH_3$), 2.20 (s, 1H, OH), 3.68 (d, 1H, $J_{\text{gem CH}_2} = 11.0$ Hz, CH₂),

3.87 (d, 1H, $J_{\text{gem CH}_2} = 11.0 \text{ Hz}$, CH₂), 4.05 (m, 2H, H-5, H-6_a), 4.23 (dd, 1H, $J_{5,6b} = 4.4, J_{6a,6b} = 12.5 \,\text{Hz}, \,\text{H}_{6b}$, 4.92 (dd, 1H, $J_{1,2} = 3.7, J_{2,3} = 9.6 \,\text{Hz}, \,\text{H}_{6a}$ 2), 5.02 (dd, 1H, $J_{3,4} = J_{4,5} = 9.6$ Hz, H–4), 5.13 (d, 1H, $J_{1,2} = 3.7$ Hz, H–1), 5.39 (dd, 1H, $J_{2,3} = J_{3,4} = 9.6$ Hz, H-3), 6.23 (d, 1H, $J_{3',4'} = 5.6$ Hz, H-4'), 7.24 (d, 1H, $J_{3',4'} = 5.6$ Hz, H-3'); ¹³C-NMR (75 MHz, CDCl₃): δ 20.6–20.7 (4 CH₃), 61.8 (C-6), 67.9 (CH₂), 68.1 (C-5), 68.3 (C-4), 70.0 (C-3), 70.4 (C-2), 96.8 (C-1), 105.6 (C-2'), 125.1 (C-4'), 148.0 (C-3'), $169.2-170.8$ (4 COCH₃), 170.1 $(C-5')$.

(2'-Hydroxy-5'-oxo-2',5'-dihydrofuran-2'-yl)methyl-2,3,4,6tetra-O-acetyl- β -p-glucopyranoside (15 β R+S)

Compound 15 β was prepared analogously to the same method than 14 α Compound 10 β (302 mg, 0.26 mmol) was irradiated as 9α leading to 15 β $(260 \text{ mg}, 0.57 \text{ mmol}, 86%)$ as a mixture of diastereoisomers $(R/S: 1/1)$. A second preparative plate chromatography (dichloromethane/ethyl acetate: 50/50) led to two fractions I and II having one of both epimers in a bigger amount. 15β (R+S): solid; mp 137°C; fraction I: ¹H-NMR (200 MHz, CDCl₃): δ 2.00–2.10 (4s, 12H, –OCOCH₃), 2.08 (s, 1H, OH); 3.78 (m, 1H, H–5), 3.91 (d, 1H, $J_{\text{gem CH}_2} = 11.2 \text{ Hz}$, CH₂), 4.08 (d, 1H, $J_{\text{gem CH}_2} = 11.2 \text{ Hz}$, CH₂), 4.16 (dd, 1H, $J_{5,6a} = 2.4$, $J_{6a,6b} = 12.5$ Hz, $H-6_a$), 4.26 (dd, 1H, $J_{5,6b} = 4.6$, $J_{6a,6b} = 12.5 \text{ Hz}, H-6_b$, 4.63 (d, 1H, $J_{1,2} = 7.9 \text{ Hz}, H-1$), 4.93 (dd, 1H, $J_{1,2} = 7.9, J_{2,3} = 9.4 \text{ Hz}, \text{ H}-2, 5.05 \text{ (dd, 1H, } J_{3,4} = J_{4,5} = 9.4 \text{ Hz}, \text{ H}-4, 5.20 \text{ Hz}$ (dd, 1H, $J_{2,3} = J_{3,4} = 9.4$ Hz, H-3), 6.15 (d, 1H, $J_{3',4'} = 5.7$ Hz, H-4'), 7.23 (d, 1H, $J_{3',4'} = 5.7$ Hz, H-3'); ¹³C-NMR (75 MHz, CDCl₃): δ 20.6–20.7 (4 CH₃), 62.0 (C–6), 68.3 (C–4), 70.4 (CH₂), 71.2 (C–2), 71.8 (C–5), 72.5 (C–3), 101.3 $(C-1)$, 105.9 $(C-2')$, 125.5 $(C-4')$, 152.4 $(C-3')$, 169.2-170.8 $(4 \quad COCH_3)$, 170 (C-5'); fraction II: ¹H NMR (200 MHz, CDCl₃): δ 2.00-2.10 (4s, 12H, $-OCOCH₃$), 2.08 (s, 1H, OH); 3.78 (m, 1H, H-5), 3.87 (d, 1H, J_{gem} $_{\text{CH}_2}$ = 11.2 Hz, CH₂), 3.94 (d, 1H, $J_{\text{gem CH}_2}$ = 11.2 Hz, CH₂), 4.16 (dd, 1H, $J_{5,6a} = 2.4, J_{6a,6b} = 12.5 \text{ Hz}, H-6_a$, 4.26 (dd, 1H, $J_{5,6b} = 4.4, J_{6a,6b} = 12.5 \text{ Hz},$ $H-6_b$, 4.68 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1), 4.96 (dd, 1H, $J_{1,2} = 7.9$, $J_{2,3} = 9.6$ Hz, H-2), 5.02 (dd, 1H, $J_{3,4} = J_{4,5} = 9.6$ Hz, H-4), 5.23 (dd, 1H, $J_{2,3} = J_{3,4} = 9.6 \text{ Hz}, \text{ H}-3$, 6.19 (d, 1H, $J_{3',4'} = 5.6 \text{ Hz}, \text{ H}-4'$), 7.27 (d, 1H, $J_{3',4'} = 5.6 \,\text{Hz}, \text{ H}-3'$); ¹³C-NMR (75 MHz, CDCl₃): $\delta 20.6-20.7$ (4 CH₃), 61.8 $(C-6)$, 68.2 $(C-4)$, 70.9 $(C-2)$, 71.8 $(C-5)$, 72 $(CH₂)$, 72.4 $(C-3)$, 100.9 $(C-1)$, 105.6 (C-2'), 125.2 (C-4'), 152.9 (C-3'), $169.2-170.8$ (4 COCH₃), 170.0 (C-5').

(2'-Acetoxy-5'-oxo-2',5'-dihydrofuran-2'-yl)methyl-2,3,4,6tetra-O-acetyl- α -D-glucopyranoside (16 α)

Dry potassium acetate (417 mg, 4.25 mmol) and acetic anhydride (2.8 mL, 29.75 mmol) were added to a cooled solution of alcohol 14α (394 mg,

0.85 mmol) in dry dichloromethane (4.5 mL). The mixture was allowed to warm to rt and stirred until TLC (petroleum ether/ethyl acetate/ethyl alcohol: 48/ 47/5) indicated completed conversion (4 hr). After filtration on a pad of Celite, the organic solution was concentrated. The crude product was chromatographed twice through silica gel (petroleum ether/ethyl acetate/ethyl alcohol: 48/47/5), and then (petroleum ether/dichloromethane/diethyl ether: 33/33/33) giving 16α (240 mg, 0.48 mmol, 56%) as mixture of epimers $(R + S)$ in a ratio of $70/30$ according to NMR data; white solid, mp $68-70^{\circ}$ C; major epimer 16α (R): ¹H-NMR (200 MHz, CDCl₃): $\delta 2.00-2.10$ (5s, 15H, CH₃), 3.83 (d, 1H, $J_{\text{gem CH}_2} = 11.1 \text{ Hz}$, CH₂), 4.03 (m, 1H, H–5), 4.11 (d, 1H, $J_{\text{gem CH}_2} = 11.1 \text{ Hz}, \text{ CH}_2$), 4.14 (dd, 1H, $J_{5,6a} = 3.9, J_{6a,6b} = 12.5 \text{ Hz}, \text{ H}-6_a$), 4.23 (dd, 1H, $J_{5,6b} = 4.3$, $J_{6a,6b} = 12.5$ Hz, $H-6_b$), 4.91 (dd, 1H, $J_{1,2} = 3.7$, $J_{2,3} = 9.8 \text{ Hz}, \text{ H}-2$), 5.07 (dd, 1H, $J_{3,4} = J_{4,5} = 9.8 \text{ Hz}, \text{ H}-4$), 5.09 (d, 1H, $J_{1,2} = 3.7$ Hz, H-1), 5.36 (dd, 1H, $J_{2,3} = J_{3,4} = 9.8$ Hz, H-3), 6.31 (d, 1H, $J_{3',4'}=6.0\,\mathrm{Hz},\ \mathrm{H-4'}$), 7.56 (d, 1H, $J_{3',4'}=6.0\,\mathrm{Hz},\ \mathrm{H-3'}$); $\mathrm{^{13}C\text{-}NMR}$ (75 MHz, CDCl₃): δ 20.5–21.3 (5 CH₃), 61.7 (C–6), 67.8 (CH₂), 68.0 (C–4), 68.7 (C–2), 70.1 (C-5), 70.6 (C-3) 97.0 (C-1), 105.7 (C-2'), 124.8 (C-4'), 150.5 (C-3'), 168.3-170.0 (5 COCH₃), 170.6 (C-5'); minor epimer 16α (S): ¹H-NMR (200 MHz, CDCl₃): δ 2.00 - 2.10 (5s, 15H, CH₃), 3.83 (d, 1H, J_{gem} $_{\text{CH}_2}$ = 11.1 Hz, CH₂), 4.03 (m, 1H, H-5), 4.08 (dd, 1H, $J_{5,6a} = 3.9$, $J_{6a,6b} = 12.5$ Hz, H-6_a), 4.11 (d, 1H, $J_{\text{gem CH}_{2}} = 11.1$ Hz, CH₂), 4.27 (dd, 1H, $J_{5,6b} = 4.3$, $J_{6a,6b} = 12.5$ Hz, H-6_b), 4.88 (dd, 1H, $J_{1,2} = 3.7$, $J_{2,3} = 9.8$ Hz, H-2), 5.07 (dd, 1H, $J_{3,4} = J_{4,5} = 9.8$ Hz, H–4), 5.08 (d, 1H, $J_{1,2} = 3.7$ Hz, H–1), 5.48 (dd, 1H, $J_{2,3} = J_{3,4} = 9.8$ Hz, H-3), 6.31 (d, 1H, $J_{3',4'} = 6.0$ Hz, H-4'), 7.58 (d, 1H, $J_{3',4'} = 6.0$ Hz, H-3'); ¹³C-NMR (75 MHz, CDCl₃): δ 20.5–21.3 (5 CH₃), 62.1 (C–6), 68.7 (CH₂), 68.4 (C–4), 69.7 (C–2), 69.9 (C–5), 70.7 (C–3) 96.4 (C-1), 105.4 (C-2'), 124.4 (C-4'), 151.1 (C-3'), $168.3-170.0$ (5 COCH₃), 170.6 $(C-5')$.

Anal. Calcd for $C_{21}H_{26}O_{14}$ (502.43) (R + S): C, 50.20; H, 5.21. Found: C, 50.52; H, 5.25.

(2'-Acetoxy-5'-oxo-2',5'-dihydrofuran-2'-yl)methyl-2,3,4,6tetra-O-acetyl- β -D-glucopyranoside (17 β)

The peracetylated compound 17β was prepared as 16α . 15β (395 mg, 0.85 mmol) led to 17β (R+S) (400 mg), the epimers of which were separated by column chromatography (petroleum ether/dichloromethane/ethyl acetate: $25/25/50$. The first eluted compound corresponded to the epimer 17 β (R) (196 mg, 0.39 mmol, 46%), while the second corresponded to 17β (S) (167 mg, 0.33 mmol, 39%); $(R/S) = 54/36$; compound 17 β (R): white solid, mp 172– 173°C; $[\alpha]_D^{20} - 111$ (c 1, CHCl₃); ¹H-NMR (200 MHz, CDCl₃): δ 2.00–2.10 (5s, 15H, CH₃), 3.74 (m, 1H, H-5), 3.94 (d, 1H, $J_{\text{gem CH}_{2}} = 10.9 \text{ Hz}$, CH₂), 4.13 (dd, 1H, $J_{5,6a} = 2.3$, $J_{6a,6b} = 12.4$ Hz, H–6_a), 4.26 (dd, 1H, $J_{5,6b} = 4.6$, $J_{6a,6b} = 12.4 \text{ Hz}, \text{ H}-6_{b}$), 4.28 (d, 1H, $J_{\text{gem CH}_{2}} = 10.9 \text{ Hz}, \text{ CH}_{2}$), 4.57 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1), 4.93 (dd, 1H, $J_{1,2} = 7.9$, $J_{2,3} = 9.3$ Hz, H-2), 5.05 (dd, 1H, $J_{3,4} = J_{4,5} = 9.3$ Hz, H-4), 5.18 (dd, 1H, $J_{2,3} = J_{3,4} = 9.3$ Hz, H-3), 6.23 (d, 1H, $J_{3',4'} = 5.7$ Hz, H-4'), 7.45 (d, 1H, $J_{3',4'} = 5.7$ Hz, H-3'); ¹³C-NMR $(75 \text{ MHz}, \text{ CDCl}_3)$: $\delta 20.5-21.3$ (5 CH₃), 61.7 (C-6), 68.2 (C-4), 68.6 (CH₂), 70.8 (C-2), 72 (C-5), 72.5 (C-3), 101.0 (C-1), 105.6 (C-2'), 124.3 (C-4'), 151.0 (C-3'), 168.1-170.1 (5 COCH₃), 170.6 (C-5'); minor epimer 17 β (S): white solid, mp 153°C; $[\alpha_{\rm D}^{20}]$ -18 (c 1, CHCl₃); ¹H-NMR (200 MHz, CDCl₃): δ 2.00–2.10 (5s, 15H, CH₃), 3.77 (m, 1H, H–5), 3.94 (d, 1H, J_{gem} $_{\mathrm{CH}_2}=10.8\,\mathrm{Hz},\ \mathrm{CH}_2),\ 4.12\,$ (dd, $\ 1\mathrm{H},\ J_{5,6\mathrm{a}}=2.0,\ J_{6\mathrm{a},6\mathrm{b}}=12.0\,\mathrm{Hz},\ \mathrm{H}-6_\mathrm{a}),\ 4.27$ (dd, 1H, $J_{5,6b} = 4.4$, $J_{6a,6b} = 12.0$ Hz, $H-6_b$), 4.28 (d, 1H, $J_{\text{gem CH}_2} = 10.8$ Hz, CH₂), 4.58 (d, 1H, $J_{1,2} = 7.6$ Hz, H-1), 4.94 (dd, 1H, $J_{1,2} = 7.6$, $J_{2,3} = 9.3$ Hz, H–2), 5.06 (dd, 1H, $J_{3,4} = J_{4,5} = 9.3$ Hz, H–4), 5.20 (dd, 1H, $J_{2,3} = J_{3,4} = 9.3 \text{ Hz}, \text{ H}-3, 6.25 \text{ (d, 1H, } J_{3',4'} = 5.7 \text{ Hz}, \text{ H}-4', 7.58 \text{ (d, 1H, }$ $J_{3',4'} = 5.7 \,\text{Hz}, \text{ H}-3'$); ¹³C-NMR (75 MHz, CDCl₃): $\delta 20.5-21.3$ (3 CH₃), 61.6 $(C-6)$, 68.1 $(C-4)$, 69.9 $(CH₂)$, 70.1 $(C-2)$, 72.1 $(C-5)$, 72.5 $(C-3)$, 101.0 $(C-$ 1), 105.4 $(C-2')$, 124.2 $(C-4')$, 151.3 $(C-3')$, 168.1-170.2 (5 $COCH₃$), 170.6 $(C-5')$.

Anal. Calcd for $C_{21}H_{26}O_{14}$ (502.43) (R): C, 50.20; H, 5.21. Found: C, 50.18; H, 5.22.

Anal. Calcd for $C_{21}H_{26}O_{14}$ (502.43) (S): C, 50.20; H, 5.21. Found: C, 50.20; H, 5.25.

(5'-Oxo-2',5'-dihydrofuran-2'-yl)methyl-2,3,4,6-tetra-Oacetyl- α -D-glucopyranoside (1 α)

Sodium borohydride (8.7 mg, 0.23 mmol) was added by little portion to a solution of 14α (106 mg, 0.23 mmol) in dry methanol (10 mL) at rt. The mixture was stirred until TLC (dichloromethane/ethyl acetate: 66/34) indicated completed conversion (1 hr) at rt. Then pH of the solution was reached up to 1 by adding aq. hydrogen chloride (5N). After stirring for 1 hr, water (10 mL) and dichloromethane (3 mL) were added. The organic phase was treated with aq. sodium hydrogenocarbonate solution, dried, and evaporated. The crude product was purified on column chromatography (dichloromethane/ethyl acetate: 66/34) giving 1α (R + S) (8.1 mg, 0.184 mmol, 80%) as a colorless oil. The reduction in the presence of cesium sulfate led to 1α (S) in a smaller amount, allowing to attribute the NMR data of each epimer. Epimer 1α (R): ¹H-NMR (200 MHz, CDCl₃): $\delta 2.00 - 2.10$ (4s, 12H, CH₃), 3.87 (dd, 1H, J_{CH_a} , $H_{2'} = 4.3$, $J_{\text{gem CH}_2} = 12.2 \text{ Hz}$, $-OCH_2$), 3.95 (m, 1H, H–5), 4.07 (dd, 1H, $J_{\text{CH}_b}, H_{2'} = 2.2, J_{\text{gem CH}_2} = 12.2 \text{ Hz}, -\text{OCH}_2$), 4.10 (dd, 1H, $J_{5,6a} = 2.2$, $J_{6a,6b} = 12.2 \text{ Hz}, H_{6a}$, 4.22 (dd, 1H, $J_{5,6b} = 4.2, J_{6a,6b} = 12.2 \text{ Hz}, H_{6b}$), 4.90 (dd, 1H, $J_{1,2} = 3.6$, $J_{2,3} = 10.0$ Hz, H-2), 5.06 (dd, 1H, $J_{3,4} =$ $J_{4,5} = 10.0$ Hz, H-4), 5.14 (d, 1H, $J_{1,2} = 3.6$ Hz, H-1), 5.21 (m, 1H, H-2'),

 5.43 (dd, 1H, $J_{2,3} = J_{3,4} = 10.0$ Hz, H-3), 6.25 (dd, 1H, $J_{2',4'} = 2.0$, $J_{3'}$ $\mathcal{A}_{\mathcal{A}'} = 5.5\,\text{Hz}, \; \text{H-4'}), \; 7.51 \; (\text{dd}, \; 1\text{H}, \; J_{2',3'}=1.5, \; J_{3',4'}=5.5\,\text{Hz}, \; \text{H-3'}); \; \text{^{13}C-NMR}$ $(75 \text{ MHz}, \text{ CDCl}_3)$: $\delta 20.7-20.8$ (4 CH₃), 61.9 (C-6), 67.5 (CH₂), 67.9 (C-5), 68.4 (C-4), 69.9 (C-3), 70.7 (C-2), 81.6 (C-2'), 96.7 (C-1), 123.5 (C-4'), 152.6 (C-3'), 170.1-170.6 (4 COCH₃), 172.2 (C-5'); epimer 1α (S): ¹H-NMR (200 MHz, CDCl₃): δ 2.00–2.10 (4s, 12H, CH₃), 3.93 (dd, 1H, $J_{\text{CH}_\text{a},\text{H2}} = 4.3$, $J_{\text{gem CH}_2} = 12.2 \text{ Hz}, -\text{OCH}_2$, 3.95(m, 1H, H–5), 4.07 (dd, 1H, $J_{\text{CH}_{\text{b}}\text{,H2'}} = 2.2$, $J_{\text{gem CH}_2} = 12.2 \text{ Hz}, -OCH_2$, 4.10 (dd, 1H, $J_{5,6b} = 2.2, J_{6a,6b} = 12.2 \text{ Hz}, H$ - (6_b) , 4.29 (dd, 1H, $J_{5,6a} = 4.2$, $J_{6a,6b} = 12.2$ Hz, H -6_a), 4.85 (dd, 1H, $J_{1,2} = 3.5$, $J_{2,3} = 10.0$ Hz, H-2), 5.06 (dd, 1H, $J_{3,4} = J_{4,5} = 10.0$ Hz, H-4), 5.07 (d, 1H, $J_{1,2} = 3.5$ Hz, H-1), 5.23 (m, 1H, H-2'), 5.41 (dd, 1H, $J_{2,3} = J_{3,4} = 10.0$ Hz, H-3), 6.23 (dd, 1H, $J_{2',4'} = 2.0$, $J_{3',4'} = 5.5$ Hz, H-4'), 7.49 (dd, 1H, $J_{2',3'} = 1.5$, $J_{3',4'} = 5.5 \,\text{Hz}, \text{ H}-3'$); ¹³C-NMR (75 MHz, CDCl₃): $\delta 20.7-20.8$ (4 CH₃), 61.8 $(C-6)$, 67.5 $(CH₂)$, 67.7 $(C-5)$, 68.5 $(C-4)$, 70.0 $(C-3)$, 70.4 $(C-2)$, 81.4 $(C 2'$), 96.6 (C-1), 123.5 (C-4'), 152.5 (C-3'), 169.6-170.4 (4 COCH₃), 172.3 $(C-5')$; a fraction of chromatography contained pure 1α (R) in very little amount; HRMS calcd for $C_{19}H_{25}O_{12}$ [M + H]⁺: 445.1346. Found: 445.1343.

The compound 1α (S) was also obtained *via* the anomerisation of 2β (S), according to Dasgupta's procedure^[20] in nitromethane as solvent and titane tetrachloride as catalyst. The conversion rate was equal to 80% but was adequate to determine the NMR data of 1α (S) obtained from the reduction of 14α .

(5'-Oxo-2',5'-dihydrofuran-2'-yl)methyl-2,3,4,6-tetra-Oacetyl- β -p-glucopyranoside (2 β)

This dihydrofuranone 2β was prepared as 1α . The same amount of 15β afforded after purification by column chromatography (dichloromethane/ ethyl acetate: 65/35) 2β (83 mg, 0.187 mmol, 81%) as a mixture of epimers $(R/S: 1/1)$ while in the presence of cerium sulfate the epimer (R) was slightly major. 2β colorless oil; epimer 2β (R): ¹H-NMR (200 MHz, CDCl₃): δ 2.00–2.10(4s, 12H, CH₃), 3.70 (m, 1H, H–5), 3.75 (dd, 1H, $J_{\text{CH}_n,H_v} = 4.6$, $J_{\text{gemCH}_2} = 11 \text{ Hz}, -OCH_2$), 4.08 (dd, 1H, $J_{\text{CH}_b,H_2} = 4.9, J_{\text{gemCH}_2} = 11.0 \text{ Hz},$ $-OCH_2$), 4.16 (dd, 1H, $J_{5,6a} = 2.2$, $J_{6a,6b} = 12.4$ Hz, H -6_a), 4.26 (dd, 1H, $J_{5,6b} = 4.2, J_{6a,6b} = 12.4 \text{ Hz}, \text{ H}-6_b$, 4.53 (d, 1H, $J_{1,2} = 7.9 \text{ Hz}, \text{ H}-1$), 5.03 (dd, 1H, $J_{1,2} = 7.9$, $J_{2,3} = 9.3$ Hz, H-2), 5.10 (m, 1H, H-2'), 5.15 (dd, 1H, $J_{3,4}=J_{4,5}=9.3\,\mathrm{Hz},\ \mathrm{H-4)},\ 5.24\ (\mathrm{dd},\ 1\mathrm{H},\ J_{2,3}=J_{3,4}=9.3\,\mathrm{Hz},\ \mathrm{H-3}),\ 6.17\ (\mathrm{dd},$ 1H, $J_{2',4'} = 2.0$, $J_{3',4'} = 5.7$ Hz, H-4'), 7.47 (dd, 1H, $J_{2',3'} = 1.5$, $J_{3',4'} = 5.7$ Hz, H-3'); ¹³C-NMR (75 MHz, CDCl₃): δ 20.7-20.8 (4 CH₃), 61.8 (C-6), 68.1 $(CH₂), 68.2 (C-4), 71.0 (C-2), 72.5 (C-5), 72.6 (C-3), 81.8 (C-2), 101.3 (C-$ 1), 123.1 (C-4'), 153.0 (C-3'), 170.2-170.6 (4 COCH₃), 172.5 (C-5'); epimer ${\bf 2} {\boldsymbol \beta}$ (S): 1 H-NMR (200 MHz, CDCl₃): $\delta 2.00{-}2.10$ (4s, 12H, CH₃), 3.70 (m, 1H, H–5), 3.81 (dd, 1H, $J_{\text{CH}_a,H_y} = 4.6$, $J_{\text{gem CH}_2} = 11.0$ Hz, $-OCH_2$), 4.08 (dd, 1H,

 $J_{\text{CH}_b},\text{H}_{2'}=4.6,~~J_{\text{gem}}~~\text{ }_{\text{CH}_2}=11.0\text{ Hz},~~-\text{OCH}_2),~~4.12~~(\text{dd},~~1\text{H},~~J_{5,6a}=2.2,$ $J_{6a,6b} = 12.4 \text{ Hz}, \text{ H}-6_{a}$, 4.23 (dd, 1H, $J_{5,6b} = 4.5, J_{6a,6b} = 12.4 \text{ Hz}, \text{ H}-6_{b}$), 4.60 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1), 5.08 (dd, 1H, $J_{1,2} = 7.9$, $J_{2,3} = 9.2$ Hz, H-2), 5.15 (dd, 1H, $J_{3,4} = J_{4,5} = 9.2$ Hz, H-4), 5.15 (m, 1H, H-2'), 5.24 (dd, 1H, $J_{2,3} = J_{3,4} = 10.0 \,\text{Hz}, \text{ H}-3$), 6.20 (dd, 1H, $J_{2',4'} = 2, \ J_{3',4'} = 5.57 \,\text{Hz}, \text{ H}-4'$), 7.49 (dd, 1H, $J_{2',3'} = 1.5$ Hz, $J_{3',4'} = 5.7$ Hz, H-3'); ¹³C-NMR (75 MHz, CDCl₃): δ 20.6–20.7 (4 CH₃), 61.7 (C–6), 68.3 (C–4), 69.3 (CH₂), 70.8 (C–2), 72.0 (C– 5), 72.5 (C-3), 81.3 (C-2'), 100.9 (C-1), 122.7 (C-4'), 153.5 (C-3'), $169.5-$ 170.1 (4 COCH₃), 172.4 (C-5[']) similar to 2β (S) NMR data.

Methyl 4-Oxo-5((2′,3′,4′,6′-tetra-O-acetyl- α -Dglucopyranosyl)oxy]pentanoate (4 α)

To a solution of 14α (460 mg, 1 mmol) in dimethylformamide (2 mL), maintained under argon, were added potassium formate (217 mg, 2.64 mmol) and palladium acetate (6 mg, 0.03 mmol). The mixture was stirred at 60° C until TLC (dichloromethane/ethyl acetate: 88/12) indicated completed conversion (2 hr 30 min). The cooled mixture was diluted with diethyl ether (20 mL). The solid was filtered off and the solvent was evaporated. To the remaining oil, dissolved in dried acetone (64 mL) and stirred at rt, were added dimethyl sulfate $(0.6$ mL) and potassium carbonate $(4 \times 0.52$ mg, each 15 min). The mixture was stirred until TLC (dichloromethane/ethyl acetate: 88/12) indicated completed conversion (4 hr). The solid was filtered off and the solvent was evaporated. The crude product was purified by column chromatography (dichloromethane/ ethyl acetate: 88/12) giving 4α (357 mg, 0.75 mmol, 75%), which was recrystallized in ethyl alcohol. 4α : white solid, mp $48^{\circ}\mathrm{C};$ $[\alpha]_{\mathrm{D}}^{20}+158$ (c 0.1, CHCl₃); $^1\mathrm{H}$ -NMR (200 MHz, CDCl₃): δ 2.00–2.10 (4s, 12H, CH₃), 2.75 (m, 4H, $J_{2a,3a}$ = $J_{2b3a} = 4.8, J_{2a,2b} = J_{3a,3b} = 12.0 \text{ Hz}, \text{ H}-2, \text{ H}-3, 3.66 \text{ (s, 3H, } -OCH_3), \text{ 4.07}$ (dd, 1H, $J_{5',6'a} = 2.2$, $J_{6'a,6'b} = 10.5$ Hz, $H-6'_a$), 4.15 (m, 2H, $H-5'$, $H-6'_b$), 4.26 (s, 2H, H-5), 4.90 (dd, 1H, $J_{1',2'} = 3.7$, $J_{2',3'} = 10.2$ Hz, H-2'), 5.07 (dd, 1H, $J_{3',4'} = J_{4',5'} = 9.6$ Hz, H-4'), 5.09 (d, 1H, $J_{1',2'} = 3.7$ Hz, H-1), 5.51 (dd, 1H, $J_{2',3'}=10.2,\,\,J_{3',4'}=9.6\,\mathrm{Hz},\,\,\,\mathrm{H-3});\,\,\,{}^{13}\mathrm{C\text{-}NMR}\,\,\,{}(75\,\mathrm{MHz},\,\,\, \mathrm{CDCl}_3);\,\,\, \delta20.6\mathrm{-}20.7$ $(4 \text{ CH}_3), 27.3 \text{ (C--2)}, 33.5 \text{ (C--3)}, 51.9 \text{ (-OCH}_3), 61.8 \text{ (C--6)}, 67.9 \text{ (C--4'), } 68.4$ $(C-5)$, 69.9 $(C-2)$, 70.5 $(C-3')$, 72.2 $(C-5)$, 96.2 $(C-1')$, 169.6-170.6 (4 COCH₃), 172.8 (C-4), 204.9 (C-1).

Anal. Calcd for $C_{20}H_{28}O_{13}$ (472.42): C, 50.42; H, 5.92. Found: C, 50.20; H, 5.85.

Methyl 4-Oxo-5((2΄,3΄,4΄,6΄-tetra-O-acetyl-β-Dglucopyranosyl)oxy]pentanoate (5 β)

The β ceto ester 5β was prepared as 4α . The same amount of 15β (460 mg, 1 mmol) gave 5β (330 mg, 0.7 mmol, 70%) as a white solid, which was

recrystallized in ethyl alcohol. 5β : mp 89°C; [α] $_{\rm D}^{20}{\rm -29}$ (c 0.8, CHCl $_{\rm 3}$); $^{\rm 1}$ H-NMR (200 MHz, CDCl₃): δ 2.00–2.10 (4s, 12H, CH₃), 2.57 (m, 2H, $J_{2a,3} = 4.3$, $J_{2b,3a} = 6.0, J_{2a,2b} = 12.0 \text{ Hz}, H-2, 2.75 \text{ (m, 2H, } J_{2,3a} = 4.3, J_{2,3b} = 6.0,$ $J_{3a,3} = 12.0 \,\text{Hz}$, H-3), 3.65 (s, 3H, $-OCH_3$), 3.73-3.78 (m, 2H, H-5', H-6'_a), 4.13 (dd, 1H, $J_{5',6'b} = 6.6$, $J_{6'a,6'b} = 12.3 \text{ Hz}$, H-6_b), 4.21 (d, 1H, $J_{5a,5b} = 12.0 \text{ Hz}, \text{ H-5a}, \text{ 4.25 (d, 1H, } J_{5a,5b} = 12.0 \text{ Hz}, \text{ H-5b}, \text{ 4.56 (d, 1H, }$ $J_{1',2'}$ = 7.6 Hz, H–1′), 5.06 (dd, 1H, $J_{1',2'}$ = 7.6, $J_{2',3'}$ = 9.3 Hz, H–2′), 5.10 (dd, 1H, $J_{3',4'} = J_{4',5'} = 9.3$ Hz, H-4'), 5.21 (dd, 1H, $J_{2',3'} = J_{3',4'} = 9.3$ Hz, H-3'); ¹³C-NMR (75 MHz, CDCl₃): δ 20.6–20.7 (4 CH₃), 27.2 (C–2), 33.7 (C–3), 51.8 $(-OCH₃), 61.7 (C–6'), 68.3 (C–4'), 71.1 (C–5'), 72.0 (C–2'), 72.6 (C–3'), 73.7$ $(C-5)$, 100.7 $(C-1')$, 169.4-170.6 (4 $COCH_3$), 173.0 $(C-4)$, 206.2 $(C-1)$.

Anal. Calcd for C₂₀H₂₈O₁₃ (472.42): C, 50.42; H, 5.92. Found: C, 50.70; H, 5.95.

ACKNOWLEDGMENT

The participation of Sudzücker AG (Mannheim, Germany) in providing HMF and isomaltulose is gratefully acknowedged.

REFERENCES

- [1] Cottier, L.; Descotes, G. 5-Hydroxymethylfurfural, syntheses and chemical transformations. Trends in Heterocycle Chem. 1991, 2, 233–248.
- [2] Neyret, C.; Cottier, L.; Descotes, G. (Beghin-Say S.A. Fr.) Preparation of 5-hydroxymethylfurfural from saccharides. Fr. 2, 663, 993, Jan. 1992; Chem. Abstr. 1992, 117, 90121b, and New process for the preparation of 5-hydroxymethylfurfural by thermal degradation of sacchardies. Fr. 2, 664, 273, Jan. 1992; Chem. Abstr. 1992, 117, 48323u.
- [3] Cottier, L.; Descotes, G.; Eymard, L.; Rapp, K. Syntheses of γ -oxo acids or γ -oxo esters by photooxygenation of furanic compounds and reduction under ultrasound: application to the synthesis of 5-amniolevulinic acid hydrochloride. Synthesis 1995 (3), 303–306.
- [4] Cottier, L.; Descotes, G.; Eymard, L.; Rapp, K. (Südzucker, Ger.) Process for the preparation of N-acylderivatives of 5-aminolevulinic acid and their hydrochlorides. Ger DE 4,228,084, Dec 1993; Chem. Abstr. 1994, 120, 191132y.
- [5] Li, R.Z.; Ji, X.J. The cytoxicity and action mechanism of ranunculin in vitro. "Yao xue xue bao." 1993, 28, 326–331; Chem. Abstr. 1993, 119, 62606m.
- [6] Li, R.Z.; Pei, H.P.; Ji, X.J. Antimutagenic activity and metabolic transformation of ranunculin by rat liver microsomes. "Yao xue xue bao." 1993, 28, 481–485; Chem. Abstr. 1993, 119, 240986v.
- [7] Lichtenthaler, F.W.; Brust, A.; Cuny, E. Sugar derived building block. Part 26. Hydrophilic pyrroles, pyridazines and diazepinones from D-fructose and isomaltulose. Green Chem. 2001, 3 (5), 201–209.
- [8] Lichtenthaler, F.W. Unsaturated O and N heterocycles from carbohydrates feedstocks. Acc. Chem. Res. 2002, 35 (9), 728–737.
- [9] Hill, R.; Heyningen, R. Ranunculin: the precursor of the vesicant substance of the buttercup. Biochem. J. 1951, 49, 332–335.
- [10] Benn, M.H.; Yelland, L.J. Ranunculin. Can. J. Chem. 1968, 46 (5), 729–732.
- [11] Boll, P.M. Naturally occuring lactones and lactames. I. Absolute configuration of ranunculin, lichesterinic acid and some lactones related to lichesterinic acid. Acta Chem. Scand. 1968, 22 (10), 3245–3250.
- [12] Cardellach, J.; Estopa, C.; Font, J.; Moreno-Mañas, M.; Ortuño, R.M.; Sacheź-Fernando, F.; Valle, S.; Vilamajo, L. Studies on structurally simple α , β –butenolides—I. New syntheses of racemic γ –hydroxymethyl– α, β –butenolides and derivatives. Tetrahedron 1982, 38 (15), 2377–2394.
- [13] Camps, P.; Cardellach, J.; Font, J.; Ortuño, R.M.; Pansati, O. Studies on structurally simple $\alpha\beta$ –butenolides—II.(-)–(S)– γ –Hydroxymethyl– α,β –butenolides and derivatives from D–ribonolactone efficient synthesis of $(-)$ –ranuculin. Tetrahedron 1982, 38 (15), 2395–2402.
- [14] Fang, Z.; Zhou, J.; Huang, L. Studies of the total synthesis of $(-)$ –ranunculin. "Yao hsueh hsueh pao." 1989, 24 (3), 182–188; Chem. Abstr. 1990, 112, 77757r.
- [15] Lichtenthaler, F.W.; Martin, D.; Weber, T.; Schiweck, H. $5-(\alpha-D-Gluco s ylox$ methyl)furfural: preparation from isomaltulose and exploration of its ensuing chemistry. Liebigs Ann. Chem. 1993 (9), 967–974.
- [16] Schmidt, R.R.; Michel, J. Facile synthesis of α –and β –O–glycosyl imidates, preparation of glycosides and disaccharides. Angew. Chem., Int. Ed. Engl. 1980, 19 (9), 731–732.
- [17] Nair, V.; Joseph, J.P. Glycosidation of silyl ethers: a novel synthesis of oligosaccharides and aryl glycosides. Heterocycles 1987, 25, 337–341.
- [18] Johary, N.S.; Owen, L.N. S–Benzyl derivatives of 2 : 3–dimercaptopropanol and 1 : 3 dimercaptopropan–2–ol. J. Chem. Soc. 1955, 1302–1305.
- [19] Kumar, V.; Amann, A.; Ourisson, G.; Luu, B. Stereospecific syntheses of 7β –and 7α –hydroxycholesterols. Synth. Commun. 1987, 17 (11), 1279–1286.
- [20] Dasgupta, F.; Garegg, P.J. Synthesis of ethyl and phenyl 1–thio–1,2–trans–D glycopyranosides from the corresponding $per-O$ –acetylated glycopyranoses having a 1,2–trans–configuration using anhydrous ferric chloride as a promoter. Acta Chem. Scand. 1989, 43 (5), 471–475.
- [21] Misiti, D.; Zappia, G.; Monache, G.D. Selective catalytic hydrogenation of γ –amino α, β –unsaturated esters in the presence of hydrogenable protecting groups. Synthesis 1999 (5), 873–877.
- [22] Arcadi, A.; Bernochi, E.; Cacchi, S.; Marinelli, F. Palladium catalysed conjugate reduction of α,β –unsaturated carbonyl compounds with potassium formate. Synlett 1991 (1), 27–28.
- [23] Yu, L.; Chen, D.; Li, J.; Wang, P.G. Preparation, characterization and synthetic uses of lanthanide (III) catalysts supported on ion exchange resins. J. Org. Chem 1997, 62, 3575–3581.
- [24] Cottier, L.; Descotes, G.; Lewkowski, J. Synthesis of furan–2,5–dicarbaldehyde by oxidation of 5–siloxymethyl–2–furfural. Synth. Commun. 1994, 24 (7), 939–944.
- [25] Bernasconi, C.; Cottier, L.; Descotes, G.; Nigay, H.; Parron, J.C.; Wiswiewski, A. Photo–oxygenation of 5–hydroxymethylfurfur–2–al. Bull. Soc. Chim. Fr., 1984 (7,8), II–323–327.